Amendments to the Specification

Please replace the paragraph beginning at page 19, line 25, with the following amended paragraph:

The partially purified enzyme preparation having α-isomaltosyl glucosaccharide-forming activity, thus obtained, was dialyzed against 10 mM sodium phosphate buffer (pH 7.0) with 1 M ammonium sulfate. The dialyzed solution was centrifuged to remove impurities, and subjected to an affinity column chromatography using 500 ml of "SEPHACRYL HR S-200", a gel commercialized by Amersham Biosciences K. K., Tokyo, Japan. The enzyme was adsorbed on "SEPHACRYL HR S-200" gel and, when eluted with a linear gradient decreasing from 1 M to 0 M of ammonium sulfate, the enzymatic activity was eluted with a linear gradient of ammonium sulfate at about 0.2 M, and fractions with the enzyme activity was collected as purified enzyme preparation. ~~The amount of enzyme activity, specific activity and yield of the α-isomaltosyl glucosaccharide-forming enzyme in each purification step are in Table2.~~

Please replace the paragraph beginning on page 23, line 3, with the following amended paragraph:

A test for investigating whether the following each saccharide can be used or not as a glucosyl donor for transferring glucosyl residue to L-ascorbic acid by α-isomaltosyl glucosaccharide-forming enzyme was carried out. A solution containing glucose, maltose, maltotriose, maltotetraose, maltopentaose, maltohexaose, maltoheptaose, isomaltose, isomaltotriose, isopanose, trehalose, kojibiose, nigerose, neotrehalose, cellobiose, gentiobiose, maltitol, maltotriitol, lactose, sucrose, erlose, selaginose, maltosylglucoside, isomaltosylglucoside, α-cyclodextrin, β-cyclodextrin, γ-cyclodextrin, amylose, amylopectin, glycogen, pullulan, or dextran was prepared. To each solution, L-ascorbic acid was further added and the concentrations of the saccharide and L-ascorbic acid were adjusted to 2% (w/v). To each substrate solution, purified α-isomaltosyl glucosaccharide-forming enzyme preparation, obtained by the method in Experiment2, was added to give 3 units/g-substrate of the amount of enzyme, and the concentration of the substrate was adjusted to 1.6% (w/v), and followed the enzyme reaction at 40°C, pH 6.0 for 20 hours. The formation of AA-2G in the reaction mixture was detected by thin-layer chromatography (abbreviated as "TLC", hereinafter) using a silica gel plate.

A mixture of n-butanol, pyridine, and water (volume ratio of 6:4:1) was used as a solvent for the development. After developing samples once on "KIESELGEL 60F_{254}", silica gel-aluminum plate (20 x 20 cm) commercialized by Merck Ltd. Japan, Tokyo, Japan, AA-2G and L-ascorbic acid were detected by irradiating ultraviolet-ray. From the results of TLC analysis, the formation of AA-2G was evaluated. ~~The results are in Table 3.~~

Please replace the paragraph beginning at page 24, line 5, with the following amended paragraph:

~~As is evident from the results in Table 3, it~~It was revealed that α-isomaltosyl glucosaccharide-forming enzyme formed AA-2G by using saccharides, having a maltose structure at their non-reducing end and the glucose polymerization degree of three or higher as glucosyl donors and transferring a glucosyl residue to L-ascorbic acid. Further, it was also revealed that α-isomaltosyl glucosaccharide-forming enzyme formed AA-2G by using saccharides, having a glucose polymerization degree of two such as maltose, kojibiose, nigerose, neotrehalose, maltotriitol, erlose as glucosyl donors.